

Available online at www.sciencerepository.org

Science Repository



Case Report and Review of the Literature

Hematopoietic Stem Cell Transplantation in Juvenile Myelomonocytic Leukemia: A Case Report and Literature Review

Malek Benakli*, Redhouane Ahmed Nacer, Farih Mehdi, Mounira Baazizi, Nadia Rahmoune, Dina Ait Ouali, Hanane Bouarab, Sara Zerkout, Fouzia Louar and Rose-Marie Hamladji

Hematology and Bone Marrow Transplant Department, Pierre and Marie Curie Center, Algiers, Algeria

ARTICLE INFO

Article history:

Received: 4 August, 2022

Accepted: 9 September, 2022

Published: 14 October, 2022

Keywords:

*Juvenile myelomonocytic leukemia
myelodysplastic/myeloproliferative
diseases
pediatric leukemia
allogeneic hematopoietic stem cell
transplantation*

ABSTRACT

Juvenile myelomonocytic leukemia (JMML) is a rare hematological malignancy of early childhood, classified by the World Health Organization as a myelodysplastic/myeloproliferative disease and is associated with a poor prognosis. Allogeneic hematopoietic stem cell transplantation is the only curative treatment. A two-year-old male child was diagnosed with JMML and was given induction chemotherapy. One year after diagnosis, the patient received allogeneic hematopoietic stem cell transplantation from an HLA sibling donor after a myeloablative conditioning regimen. The patient remained free of disease after 5 years of follow-up, healthy, with complete clinical, immunologic and hematologic recovery, without signs of JMML. Transplantation is the only modality to achieve a cure in JMML patients. The most widely practiced approach is the use of bone marrow or peripheral blood stem cells after a myeloablative conditioning regimen. Post-transplant monitoring chimerism can help identify the patients who are at risk of relapse.

© 2022 Malek Benakli. Hosting by Science Repository.

Introduction

Juvenile myelomonocytic leukemia (JMML) is a rare hematological malignancy of early childhood representing 2 to 3% of all pediatric leukemia cases; classified by the World Health Organization (WHO) as a myelodysplastic/myeloproliferative disease and is associated with poor prognosis. Allogeneic hematopoietic stem cell transplantation (allo-HSCT) is the only curative treatment [1-3].

Case Presentation

A two-year-old male child presented with pallor, fever, skin rash and pain in the abdomen for three weeks. The patient had no similar complaints in the past. The birth history was normal. The patient was fully vaccinated and had normal developmental milestones. He had cervical lymph nodes of size ranging between 1 to 2 cm. The spleen and liver were palpable 4 and 3 cm below the costal margin, respectively. CT-scan revealed abdominal lymphadenopathy and hepatosplenomegaly. No significant abnormality was seen with the X-

ray thoracic chest. Complete blood cell count showed leukocytosis ($31.8 \times 10^9/L$), anemia (8.9 g/dL), normal platelet counts ($368 \times 10^9/L$) and absolute monocyte count ($7.9 \times 10^9/L$). Morphologic evaluation of peripheral blood smear noted a myeloid leukemia with 10% of myelocytes, metamyelocytes, and erythroblasts. Bone marrow (BM) aspiration showed hypercellularity with dysgranulopoiesis and 4% blasts. No BCR-ABL fusion transcript was found and karyotype on BM cells shows trisomy 8. The synthesis of fetal haemoglobin (HbF) was 0.2%. Abnormal JMML-related mutations were not done. He was diagnosed as JMML and was given induction chemotherapy with cytarabine (25 mg/m²/d) and Mercaptopurine (50 mg/m²/d), 5 days/month, 8 courses, with resulted in regression of hepatosplenomegaly and normalization of his total and differential counts.

One year after diagnosis, the child received peripheral blood stem cell (PBSC) as a graft (CD 34 cell counts: $8.7 \times 10^6/kg$) from an HLA sibling donor after a conditioning regimen associated with Busilvex and Cyclophosphamide (200 mg/kg) and GVHD prophylaxis with cyclosporin and methotrexate. The time required for achieving a

*Correspondence to: Malek Benakli, Hematology and Bone Marrow Transplant Department, Pierre and Marie Curie Center, Avenue Battandier, Algiers, Algeria, ORCID: 0000-0001-8444-890X; Tel: +21321237025; Fax: +21321237025; E-mail: malekbenakli@gmail.com

granulocyte count greater than $0.5 \times 10^9/l$ was 22 days. No complications noted: severe infection, cytomegalovirus (CMV) reactivation, acute or chronic graft versus host disease (GVHD). Chimerism was fully donor (> 95% donor) at days +30, +100, +180 and +365 posttransplant. The patient remained free of disease after 5 years of follow-up, healthy, with normal growth, complete clinical, immunologic and hematologic recovery, without signs of JMML.

Discussion

JMML has a peak incidence at the age of 2 years and patients present with various symptoms that could also be associated with infection. The

diagnostic criteria of JMML are presented in (Table 1). The common clinical symptoms include fever, coughing, lymphadenopathy, hepatosplenomegaly, cutaneous lesions and skin bleeding. Usually, a blood examination shows high white blood cell counts, anaemia, thrombocytopenia, monocytosis and elevated haemoglobin F. Myeloid precursors are found in the morphologic evaluation of peripheral blood smear and the percentage of blasts in peripheral blood and bone marrow is lower than 20%. Monosomy of chromosome 7 is the most frequent karyotypic abnormality and more than 85% of the patients have a mutation in *K-RAS*, *N-RAS*, *PTPN-11*, *CBL* or *NF1*, causing aberrant activation of the RAS signalling pathway and GM-CSF-hypersensitivity [1, 4, 5].

Table 1: The diagnostic criteria for JMML (2016 WHO classification).

<p>I. Clinical and hematologic features (all 4 features mandatory)</p> <ul style="list-style-type: none"> - Peripheral blood monocyte count $\geq 1.10^9/L$ - Blast percentage in peripheral blood and bone marrow < 20% - Splenomegaly - Absence of Philadelphia chromosome (BCR/ABL rearrangement)
<p>II. Oncogenetic studies (1 finding is sufficient)</p> <ul style="list-style-type: none"> - Somatic mutation in <i>PTPN11</i> or <i>K-RAS</i> or <i>N-RAS</i> * - Clinical diagnosis of <i>NF-1</i> or germline <i>NF1</i> mutation - Germline <i>CBL</i> mutation and loss of heterozygosity of <i>CBL</i>**
<p>III. For patients without any oncogenetic parameter, besides clinical and hematologic features listed under I, the following criteria must be met:</p> <ul style="list-style-type: none"> - Monosomy 7 or any other chromosomal abnormality or at least 2 of the following criteria: - HbF increased for age - Myeloid or erythroid precursors on peripheral blood smear - Spontaneous growth or GM-CSF hypersensitivity in colony assay - Hyperphosphorylation of <i>STAT5</i>

WHO: World Health Organization. * Germ line mutations (indicative of Noonan syndrome) need to be excluded.

** Occasional cases may harbor heterozygous splice site mutations. *NF-1*: Neurofibromin-1; *CBL*: Casitas B-lineage lymphoma; *PTPN11*: Protein Tyrosine Phosphate Non-Receptor type; *K-RAS*: Kristen Rat Sarcoma; *N-RAS*: Neuroblastoma Rat Sarcoma.

In the majority of cases, JMML is a fatal disorder in children who do not receive transplantation. Allo-HSCT is the only curative modality in JMML. It has been recognized that patients with germline *CBL* and acquired loss of heterozygosity or biallelic *CBL* mutations may not require HSCT. In contrast, patients with JMML and *NF1*, somatic *PTPN11* or *KRAS* mutation, the majority of somatic *NRAS* and those without known molecular lesions have a clear indication for HSCT [3, 6, 7]. Stieglitz *et al.* demonstrate that patients who harbor two or more somatic alterations at diagnosis had significantly worse EFS and OS compared to those with one or fewer events [8]. Therapeutic considerations range from observation to allo-HSCT, depending on the genetic subtype. Initial diagnosis at the age of > 2 years old, platelet count $\leq 33 \times 10^9/L$, Haemoglobin F level > 10%, chromosome 7, *PTPN11* mutations and combined gene mutations have been recognized as the main predictors of short survival in JMML [4, 8, 9].

The standard conditioning regimen for JMML, as originally proposed by the European Working Group of Myelodysplastic Syndromes (EWOG-MDS) in childhood, is the Busulfan-Cyclophosphamide-Melphalan (BU-CY-MEL) regimen who achieved a 52% rate of long-term disease-free survival [10]. Myeloablative conditioning regimen (MAC) with Total Body irradiation (TBI) has no benefit and suggests that among

regimens without TBI, BU-MEL with Fludarabine (FLU) could be a standard conditioning regimen in addition to the conventional BU-CY-MEL. A randomized study that compared the potentially less toxic FLU-BU with the standard BU-CY-MEL regimen was quickly closed due to the high incidence of relapse and the extremely low probability of EFS in the FLU-BU arm. One report of 10 children given orally BU, FLU (120 mg/m^2), and MEL ($180\text{-}210 \text{ mg/m}^2$) demonstrated promising results, with 7 patients in remission > 2 years since HSCT. Martenson *et al.* compared two different conditioning regimens and concluded that the addition of MEL to the BU-CY conditioning regimen resulted in severe gastrointestinal complications and did not improve overall survival. Like our patient, the most widely practiced approach is the use of BM or PBSC after MAC with alkylating drugs which appears potentially capable of eradicating stem cell disorders, such as JMML [3,10-18].

Retrospectives studies indicate that an unrelated donor (UD) offers minimal or no significant disadvantage compared with using an HLA-identical sibling and using an umbilical cord blood transplant (UCBT) have similar outcomes to that reported with different sources of HSC. A recent study suggested similar rates of acute and chronic GVHD and non-relapse mortality in mismatched haploidentical donor transplants compared to matched donors [5,18].

GVHD prophylaxis consisted of association of Cyclosporin and Methotrexate in our patient. This procedure is most commonly employed. The risk of transplant-related mortality (TRM) is approximately 13% and disease recurrence remains the major cause of treatment failure, with a higher probability of relapse (30% to 60%) within the first year [5, 12]. Chronic GVHD is an independent factor associated with a lower incidence of relapse and a higher probability of survival, suggesting the importance of graft versus leukemia (GVL) effects in JMML following HSCT [9, 11, 16-19]. Furthermore, grade 2-3 acute GVHD was associated with decreased incidence of relapse in the EUROCORD-CIBMTR (Center for International Blood and Marrow Transplantation Research) study [5].

Our patient is considered in complete remission according to the criteria proposed by Niemeyer *et al.* The remission criteria include the results of chimerism analyses. Patients who achieve neutrophil engraftment and complete donor chimerism with the disappearance of acquired cytogenetic and molecular abnormalities are considered to have a complete remission of JMML [20]. In several cases of mixed chimerism (MC), withdrawal of immunosuppressive therapy induced complete chimerism after allo-HSCT [19]. In the absence of acute GVHD, prophylaxis should be discontinued between day 60 and day 90. Donor lymphocyte infusion (DLI) is not effective for patients with MC or relapse, second allo-HSCT from either the same or different donor, should be considered [3, 5, 10, 19-22]. The clinical activity and acceptable toxicity of Azacitidine (32 mg/m²/day for 5 days, every 4 weeks), provide a rationale for its use in cases where allo-HSCT has failed, as a bridging strategy to second transplantation [22, 23]. Age > 4 years, BM blasts > 20% have been found to be associated with an increased risk of relapse [10].

A variety of pre-transplant treatments have been employed to control symptoms of JMML. Low-dose intravenous cytarabine alone or in combination with 6-mercaptopurine, and high-dose cytarabine have been used. No standard chemotherapy protocol has proven to have an impact on relapse incidence after transplantation but the favourable toxicity of azacitidine (75-100 mg/m² on 5 to 7 days, repeated every 4 weeks) and its cytoreductive potential make it an attractive option prior to allo-HSCT [12, 24-26]. Patients with JMML who transform into acute myeloid leukemia (AML) generally have dismal outcomes following HSCT [10, 27]. An EWOG-MDS/EBMT trial reported no significant differences in terms of EFS (52% vs. 50%), relapse incidence (35% vs. 38%), or TRM (13% vs. 13%) between AML-type chemotherapy and less intensive treatment. The same group indicates that spleen size at the time of transplantation and splenectomy prior to transplantation does not appear to have an impact on outcome [10]. A recent study noted better post-HSCT outcome of patients who experienced molecular response to pretransplant treatment [28].

Conclusion

Allo-HSCT is the only modality to achieve a cure in JMML patients. Azacitidine is a promising agent in recent clinical trials to achieve hematological and molecular responses in some patients with the aim of reducing the burden of disease before HSCT. The most widely practiced approach is the use of BM or PBSC after high-intensity MAC. Post-

transplant monitoring chimerism can help identify the patients who are at risk of relapse.

Conflicts of Interest

None.

REFERENCES

- Arber DA, Orazi A, Hasserjian R, Thiele J, Borowitz MJ et al. (2016) The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood* 127: 2391-2405. [[Crossref](#)]
- Patnaik MM, Tefferi A (2020) Chronic Myelomonocytic leukemia: 2020 update on diagnosis, risk stratification and management. *Am J Hematol* 95: 97-115. [[Crossref](#)]
- Locatelli F, Niemeyer CM (2015) How I treat juvenile myelomonocytic leukemia. *Blood* 125: 1083-1090. [[Crossref](#)]
- Niemeyer CM, Arico M, Basso G, Biondi A, Rajnoldi AC et al. (1997) Chronic myelomonocytic leukemia in childhood: a retrospective analysis of 110 cases. European Working Group on Myelodysplastic Syndromes in Childhood (EWOG-MDS). *Blood* 89: 3534-3543. [[Crossref](#)]
- Locatelli F, Crotta A, Ruggeri A, Eapen M, Wagner JE et al. (2013) Analysis of risk factors influencing outcomes after cord blood transplantation in children with juvenile myelomonocytic leukemia: a EUROCORD, EBMT, EWOG-MDS, CIBMTR study. *Blood* 122: 2135-2141. [[Crossref](#)]
- Matsuda K, Yoshida N, Miura S, Nakazawa Y, Sakashita K et al. (2012) Long-term haematological improvement after non-intensive or no chemotherapy in juvenile myelomonocytic leukemia and poor correlation with adult myelodysplasia spliceosome-related mutations. *Br J Haematol* 157: 647-650. [[Crossref](#)]
- Mayerhofer C, Niemeyer CM, Flotho C (2021) Current Treatment of Juvenile Myelomonocytic Leukemia. *J Clin Med* 10: 3084. [[Crossref](#)]
- Stieglitz E, Taylor-Weiner AN, Chang TY, Gelston LC, Wang YD et al. (2015) The genomic landscape of juvenile myelomonocytic leukemia. *Nat Genet* 47: 1326-1333. [[Crossref](#)]
- Niemeyer CM, Flotho C (2015) Juvenile myelomonocytic leukemia: who's the driver at the wheel? *Blood* 133: 1060-1070. [[Crossref](#)]
- Locatelli F, Nollke P, Zecca M, Korthof E, Lanino E et al. (2005) European Working Group on Childhood MDS; European Blood and Marrow Transplantation Group. Hematopoietic stem cell transplantation (HSCT) in children with juvenile myelomonocytic leukemia (JMML): results of the EWOG-MDS/EBMT trial. *Blood* 105: 410-419. [[Crossref](#)]
- Yoshida N, Sakaguchi H, Yabe M, Hasegawa D, Hama A et al. (2020) Clinical Outcomes after Allogeneic Hematopoietic Stem Cell Transplantation in Children with Juvenile Myelomonocytic Leukemia: A Report from the Japan Society for Hematopoietic Cell Transplantation. *Biol Blood Marrow Transplant* 26: 902-910. [[Crossref](#)]
- Dvorak CC, Satwani P, Stieglitz E, Cairo MS, Dang H et al. (2018) Disease burden and conditioning regimens in ASCT1221, a randomized phase II trial in children with juvenile myelomonocytic

- leukemia: A Children's Oncology Group study. *Pediatr Blood Cancer* 65: e27034. [[Crossref](#)]
13. Yabe M, Sako M, Yabe H, Osugi Y, Kurosawa H et al. (2008) A conditioning regimen of busulfan, fludarabine, and melphalan for allogeneic stem cell transplantation in children with juvenile myelomonocytic leukemia. *Pediatr Transplant* 12: 862-867. [[Crossref](#)]
 14. Martenson T, Priftakis P, Casswall T, Ringdén O, Mattsson J et al. (2013) Increased risk of gastrointestinal acute GVHD following the addition of melphalan to busulfan/cyclophosphamide conditioning. *Pediatr Transplant* 17: 285-293. [[Crossref](#)]
 15. Dvorak CC, Loh ML (2014) Juvenile myelomonocytic leukemia: Molecular pathogenesis informs current approaches to therapy and hematopoietic cell transplantation. *Front Pediatr* 2: 25. [[Crossref](#)]
 16. Lin YC, Luo CJ, Miao Y, Wang JM, Luo CY et al. (2021) Human leukocyte antigen disparities reduce relapse after hematopoietic stem cell transplantation in children with juvenile myelomonocytic leukemia: A single-center retrospective study from China. *Pediatr Transplant* 25: e13825. [[Crossref](#)]
 17. Yabe M, Ohtsuka Y, Watanabe K, Inagaki J, Yoshida N et al. (2015) Transplantation for juvenile myelomonocytic leukemia: a retrospective study of 30 children treated with a regimen of busulfan, fludarabine, and melphalan. *Int J Hematol* 101: 184-190. [[Crossref](#)]
 18. Smith FO, King R, Nelson G, Wagner JE, Robertson KA et al. (2002) Unrelated donor bone marrow transplantation for children with juvenile myelomonocytic leukaemia. *Br J Haematol* 116: 716-724. [[Crossref](#)]
 19. Inagaki J, Fukano R, Nishikawa T, Nakashima K, Sawa D et al. (2013) Outcomes of immunological interventions for mixed chimerism following allogeneic stem cell transplantation in children with juvenile myelomonocytic leukemia. *Pediatr Blood Cancer* 60: 116-120. [[Crossref](#)]
 20. Niemeyer CM, Loh ML, Cseh A, Cooper T, Dvorak CC et al. (2015) Criteria for evaluating response and outcome in clinical trials for children with juvenile myelomonocytic leukemia. *Haematologica* 100: 17-22. [[Crossref](#)]
 21. Yoshimi A, Niemeyer CM, Bohmer V, Duffner U, Strahm B et al. (2005) Chimerism analyses and subsequent immunological intervention after stem cell transplantation in patients with juvenile myelomonocytic leukaemia. *Br J Haematol* 129: 542-549. [[Crossref](#)]
 22. Yoshimi A, Mohamed M, Bierings M, Urban C, Korthof E et al. (2007) Second allogeneic hematopoietic stem cell transplantation (HSCT) results in outcome similar to that of first HSCT for patients with juvenile myelomonocytic leukemia. *Leukemia* 21: 556-560. [[Crossref](#)]
 23. Flotho C, Sommer S, Lübbert M (2018) DNA-hypomethylating agents as epigenetic therapy before and after allogeneic hematopoietic stem cell transplantation in myelodysplastic syndromes and juvenile myelomonocytic leukemia. *Semin Cancer Biol* 51: 68-79. [[Crossref](#)]
 24. Niemeyer MC, Flotho C, Lipka DB, Starý J, Rössig C et al. (2021) Response to upfront azacitidine in juvenile myelomonocytic leukemia in the AZA-JMML-001 trial. *Blood Adv* 5: 2901-2908. [[Crossref](#)]
 25. Cseh A, Niemeyer CM, Yoshimi A, Dworzak M, Hasle H et al. (2015) Bridging to transplant with azacitidine in juvenile myelomonocytic leukemia: a retrospective analysis of the EWOG-MDS study group. *Blood* 125: 2311-2313. [[Crossref](#)]
 26. Marcu A, Colita A, Radu LE, Jercan CG, Bica AM et al. (2020) Single-Center Experience With Epigenetic Treatment for Juvenile Myelomonocytic Leukemia. *Front Oncol* 10: 484. [[Crossref](#)]
 27. Honda Y, Tsuchida M, Zaïke Y, Masunaga A, Yoshimi A et al. (2014) Clinical characteristics of 15 children with juvenile myelomonocytic leukaemia who developed blast crisis: MDS Committee of Japanese Society of Paediatric Haematology/Oncology. *Br J Haematol* 165: 682-687. [[Crossref](#)]
 28. Hecht A, Meyer J, Chehab FF, White KL, Magruder K et al. (2019) Molecular assessment of pretransplant chemotherapy in the treatment of juvenile myelomonocytic leukemia. *Pediatric Blood Cancer* 66: e27948. [[Crossref](#)]