

Available online at www.sciencerepository.org

Science Repository



Review Article

Tissue Infiltrating Immune Cells and Endometrial Cancer Prognosis

Maureen L. Drakes^{1*}, Swati Mehrotra², Ronald K. Potkul³, Cheryl M. Czerlanis¹ and Patrick J. Stiff¹

¹Department of Medicine, Cardinal Bernardin Cancer Center, Loyola University Chicago, Chicago, Illinois, USA

²Department of Pathology and Laboratory Medicine, Loyola University Chicago, Chicago, Illinois, USA

³Department of Obstetrics and Gynecology, Loyola University Chicago, Chicago, Illinois, USA

ARTICLE INFO

Article history:

Received: 5 March, 2021

Accepted: 23 March, 2021

Published: 19 April, 2021

Keywords:

Endometrial cancer

disease outcome

immune cells

T cell subsets

macrophages

survival

ABSTRACT

The number of patients diagnosed with endometrial cancer surpasses that of any other gynaecological cancer. This disease is usually detected early after disease onset and with current therapy 80 percent of patients with early-stage disease reach a five-year survival milestone. However, patients with advanced or recurrent disease have a grim outcome and the five-year survival rate for these patients is only about 16 percent. In several cancer types there is accumulating evidence that immune cells play a crucial role in the initiation, progression and outcome of disease. In order to provide novel and effective immunotherapeutic treatments for advanced disease endometrial cancer, an understanding of the relevance of immune cells needs to be addressed. This review briefly discusses current knowledge in the area of immune cells and how they may alter the course of endometrial cancer, as well as the implications of these cells for novel therapy and outcome.

© 2021 Maureen L. Drakes. Hosting by Science Repository.

Introduction

Endometrial cancer (uterine corpus endometrial cancer; EC) is the predominantly occurring female gynaecologic cancer in the Western world. In the United States it is expected that there will be 66,570 new cases of EC in 2021, and about 12,940 deaths from this disease [1]. Risk factors for this disease include co-morbidities such as obesity, hypertension and diabetes [2, 3]. Additionally, EC is primarily found in postmenopausal women and in patients who are advanced in age. Conventional and successful treatment for early-stage EC consists of surgery, radiation and/or chemotherapy. This results in more than 80% of patients presented with early-stage EC reaching a 5-year overall survival rate. On the contrary, a small percentage of EC patients are usually diagnosed at the stage of advanced /recurrent disease, and such patients have a five-year survival rate as low as 16% [1]. There is potential for this poor outcome to be improved by the use of conventional therapy combined with targeted therapy or immunotherapy [4].

EC has been classified by different characterization standards, two of which we will discuss here. This disease has been grouped by the

International Federation of Gynaecology and Obstetrics (FIGO) as type 1 and type 2. Type 1 EC consists of FIGO grade 1 and grade 2 EC and comprise 80% of EC cases. Type 2 EC consists of 10-20% EC and include FIGO grade 3 more aggressive disease [5, 6]. Like most histological classifications of cancer, type 1 and type 2 EC may sometimes overlap and this classification system may not be as objective as needed. The Cancer Genome Atlas classification of EC has proved to be a good indicator of EC prognosis [7]. The four molecular groupings of this characterization are: Polymerase ε (POLE)-mutant ultramutated, microsatellite instability-high (MSI-H, hypermutated), copy number low and copy number high [7]. The MSI-H hypermutated group has the highest number of MMR defects and is most responsive to immunotherapeutic agents [7, 8]. The MMR pathway acts to repair single strand breaks, mispairings, as well as small insertions or deletions which occur during DNA replication. Germline MMR deficiencies may be associated with Lynch syndrome, which affects between 2 to 6 percent of endometrial cancer patients; however, most of the MMR pathways deficiencies are due to somatic mutations [9-11].

Much emphasis has been placed on the molecular groupings of EC as prime predictors of disease prognosis. Yet, it is important to understand

*Correspondence to: Maureen L. Drakes, Ph.D., Department of Medicine, Cardinal Bernardin Cancer Center, Loyola University Chicago, Building 112, 2160 South First Avenue, Maywood, 60153, Chicago, Illinois, USA; Tel: 7083273125; E-mail: mdrakes@luc.edu

the immune nature of the EC tumor microenvironment (TME) and how this cellular environment can regulate disease, so that we can better determine patients who will be most responsive to novel targeted and immune therapies. The immunosuppressive EC TME consists of tumor cells, immune cells, fibroblasts, cytokines, chemokines and other soluble molecules, cell surface receptors and other molecular targets. Even though the sum total of these parameters predisposes to an immune-suppressive environment, which fosters tumor immune evasion, immune cell subsets in the TME can fill dichotomous roles and induce pro-tumor mechanisms or anti-tumor immunity. This review provides an understanding of the primary cell types in the EC TME and briefly discusses how novel treatments may potentially modulate these cells to improve survival.

Opposing Roles of T Cell Subsets in the Regulation of Endometrial Cancer

T cells play a critical role in most of the cancer types as they are the ultimate effector mechanism in the response to therapy [12]. CD8+ T cells activate tumor cell killing by a cytotoxic T cell (CTL) dependent apoptosis mechanism [13, 14]. The beneficial role of CD4+ and CD8+ T cells in the TME on survival has been documented for several cancer types, however T cells in the EC TME has been less studied than that of most malignancies [15, 16]. In a study of 35 malignant and 23 non-malignant endometrial samples and peripheral blood samples, it was

found that the numbers of CD8+ T cells were lower in the endometrium of EC patients than in corresponding controls. These workers also found that CD8 expression was downregulated in EC [17]. It has also been reported that in EC, increased CD8+ T cells at the tumor invasive margin were found in tumors of low grade and was associated with prolonged overall survival (O/S) [18]. Other groups found that the higher CD8+ T cell infiltrate was correlated with prolonged survival in EC patients (Figure 1) [19, 20]. The molecular characterization groups POLE-ultramutated and MSI/ MMR deficiency (MMRd) tumors often have high CD3+ and high cytotoxic CD8+ T cells, and generally have the best outcome of the four EC groups [21-23].

In a meta-analysis of 17 studies, subgroup analysis according to the localization of CD8+ T cells, showed that patients with a high density of CD8+ T cells in the intraepithelial region of the tumor were more likely to have a good progression free survival (PF/S) than those with a high density of CD8+ T cells in the tumor stroma [24]. In the two studies of this meta-analysis where CD45RO+ memory T cells were studied there was a strong correlation between a high density of CD45RO+ T cells and O/S [24]. CD45RO+ T cells are associated with improved survival in several tumors [25, 26]. The purported role of CD8+ T cells on EC outcome is encouraging as it strongly suggests that this cancer may be highly amenable to immunotherapy by the manipulation of CD8+ cytotoxicity potential.

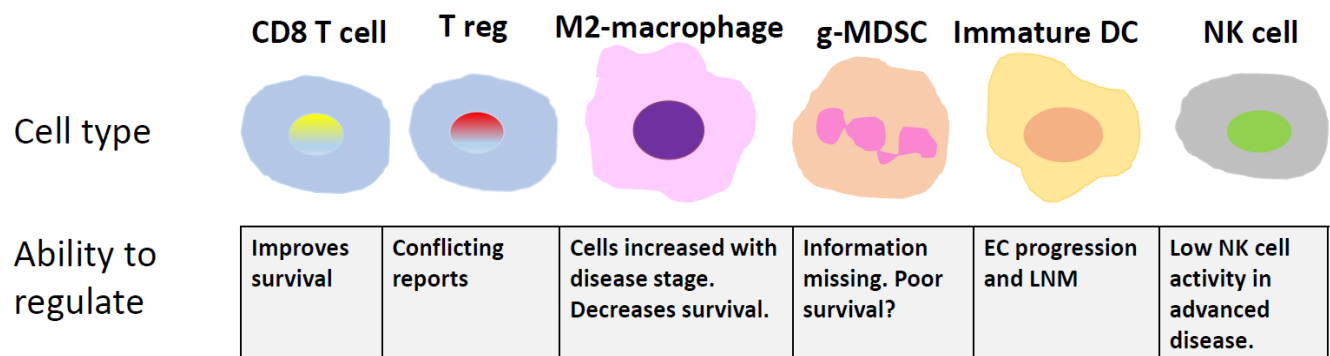


Figure 1: Primary immune modulatory cells in the endometrial cancer tumor microenvironment. The tumor microenvironment of EC consists of a variety of cell types, many of which favour disease progression and poor survival.

EC: Endometrial Cancer; DC: Dendritic Cell; g-MDSC: granulocytic Myeloid Derived Suppressor Cell; LNM: Lymph Node Metastasis; NK cell: Natural Killer cell; T reg cell: T regulatory cell.

The CD4+CD25+FoxP3+ T cell (T regs) subset has often been referred to as cells which reduce anti-tumor responses and contribute to poor survival [27]. In a study cohort of 57 EC patients, CD4+CD25+ T cells were elevated in tumor infiltrating lymphocytes (TILs) in comparison with their presence in peripheral blood lymphocytes (PBLs). This parameter had a strong positive correlation with high tumor grade, myometrium invasion, tumor stage, LNM and in general poor prognosis [28, 29]. Of note, T regs in the EC TME, but not in the PBLs expressed cytotoxic molecules granzyme B and perforin, molecules which are not usually expressed in T regs. Others have shown that T regs from the tumor environment could induce natural killer (NK) and CD8+ T cell death in a granzyme B and perforin-dependent fashion [30]. Granzyme B and perforin are both important for the ability of NK cells and CD8+ T cells to perform the function of killing their targets. Therefore, regulatory T cells may use the perforin-granzyme pathway as a

mechanism to suppress the function of immune cells through suppression of NK cells and CD8+ T cells, preventing them from clearing tumors [30]. It is reported that activated murine regulatory T cells suppressed CD4+ CD25+ T effector cells by a granzyme B-dependent mechanism [29, 31]. It remains to be investigated whether the secretion of granzyme B and perforin in the EC TME by T regs is a relevant mechanism of suppressing NK cell and/or CD8+ T cell function in this disease.

In contrast to the above reports, others have published different relationships concerning FoxP3 T regs and EC prognosis. For example, analysis of 4 studies in which FoxP3 T cells were investigated as part of a meta-analysis, concluded that FoxP3 T regs were not significantly associated with EC prognosis [24]. Another study reported that a higher fraction of infiltrating T regs was associated with better survival in EC

patients [20]. This latter finding is similar to some reports in pancreatic cancer, colorectal, bladder and esophageal cancers [20, 32-34]. Indeed, in pancreatic cancer, Zhang and colleagues found that Tregs depletion resulted in tumor progression due to a reprogramming of the fibroblast population with loss of tumor-restraining, smooth muscle actin-expressing fibroblasts, as well as other critical immune-suppressive events [34]. Overall, these studies indicate the complexity of understanding the TME of each cancer to predict how different cell subsets may influence survival.

In EC, other critical immune regulators expressed on T cells include immune checkpoint molecules such as cytotoxic T lymphocyte associated protein-4 (CTLA-4) and programmed death-1 (PD-1), among others, which contribute to T cell exhaustion and pro-tumor immune responses [35-39]. As we shall discuss in the subsequent text, therapy blocking the ligation of PD-1 to PD-L1 shows great promise in improving survival predominantly among MMRd and MSI-H EC patients [40, 41].

Messages from Macrophages in the Endometrial Cancer Tumor Microenvironment

Diverse myeloid cell subsets have been characterized in the TME of cancers with macrophages comprising about 10 percent of the cell population. Tumor associated macrophages (TAMs) may consist of the M1 (induce anti-tumor immunity) or M2 (pro-tumor) immunosuppressive polarization macrophages or myeloid derived suppressor cells [42-45]. Th1 cytokines such as IL-12 and IL-18 or Toll-like receptors induce M1 cells. These cells are efficient in killing tumor cells via secreting reactive oxygen or nitrogen species (ROS/ RNS), cytokines IL-6, IL-1 β , or tumor necrosis factor- α (TNF- α) [46]. M2 macrophage polarization is favoured by cytokines IL-4, IL-10, IL-13 and transforming growth factor- β (TGF- β), which predispose to tumor development [46].

In EC, like most cancers, tumor associated macrophages (TAMS) generally favour an immunosuppressive M2 pro-tumor environment and more severe disease [47, 48]. CD68+ macrophages have been identified in high density in the epithelial and stromal areas of EC, when compared with benign tumors [49, 50]. In another study, high numbers of CD68+ macrophages in the intra-tumoral border was associated with worse PF/S and O/S than in patients with low macrophage frequency [51]. Furthermore, in a cohort of type I EC patients, a positive correlation was shown between the frequency of TAMS, and a higher tumor grade, more advanced disease and lymph node metastasis (LNM) [52].

Natural suppressor myeloid cells were described in tumor bearing mice as early as 1964 [53, 54]. MDSC inhibit the proliferation of T cells and NK cells, stimulate tumor cell proliferation, cancer metastasis and angiogenesis [55-58]. MDSC are renowned for their involvement in cancer. These cells have been extensively studied in several cancer types and attributed to play a prime role in disease progression and resistance to conventional and novel therapies [58, 59]. Granulocytic MDSC (g-MDSC) have morphological features of granulocytes, and monocytic MDSC (m-MDSC) look like monocytes. In EC patients, however, these cells have not been well studied. In this disease it is reported that g-MDSC (CD11b+CD33+CD15+CD14-) are the primary occurring subset

in contrast to m-MDSC (CD11b+CD33+CD14+CD15-) [60, 61]. Many of the intricate networks controlling MDSC immune down-regulatory functions are mediated by arginase-1 (Arg-1), nitric oxide (NO) and signal transducer and activator of transcription-3 (STAT-3) [62-64].

Because TAMs and MDSC in particular are major contributors to cancer progression, targeting of TAMS is an excellent strategy for advanced/metastatic EC management [55, 60]. There are ongoing clinical trials targeting these cells in malignancies such as lung cancer, head and neck cancer and pancreatic cancer, but clinical trials are urgently needed in this treatment direction for other diseases such as EC [60]. Agents used to inhibit TAMs may decrease macrophage differentiation, prevent TAM recruitment or increase the efficacy of immune checkpoint blockade. CCL2 and CCL5 recruit macrophages, and pharmacologic inhibitors which block these chemokines limit TAM accumulation. Trabectedin reduces TAM in the TME, and Bevacizumab neutralizes vascular endothelial growth factor (VEGF) which is secreted by TAMS [65]. Several clinical trials blocking TAM parameters in single and combination studies in cancers are described elsewhere [60, 66].

Dendritic Cells and Endometrial Cancer

Dendritic cells (DC) are superb antigen presenting cells (APC) which capture, process and present antigen as peptides to the immune system [67]. These cells play critical roles in both health and disease [68, 69]. They are a heterogeneous group of myeloid cells which infiltrate tumors and present tumor derived antigen to antigen-specific T cells, priming these cells and eliciting anti-tumor immunity. However, the anti-tumor function of DC can be diminished by suppressive mechanisms present in the TME [70, 71]. Additionally, immature DC can be recruited to tumors and contribute to tumor progression. The expression of DC markers S100 and HLA-DR were found to correlate with EC progression and LNM [72]. Abnormal DC function is associated with EC progression. Co-stimulatory molecules CD80, CD86 and CD40 were significantly more highly expressed on DC in normal human endometrium, in comparison with the expression of these markers on DC in endometrial adenocarcinoma [73]. In the TME diminished DC co-stimulatory molecules can reduce the potency of immune responses and create an immune-suppressive environment. Several therapeutic inventions have been employed to enhance DC function in various cancer types, but to date the success of this therapy in most cancers needs to be improved to prolong survival in a significant percentage of patients [74].

Impaired Natural Killer Cell Activity in Endometrial Cancer Patients

NK cells are effector cells which bestow cytotoxic anti-tumor immunity and function in anti-viral immune responses. There is generally a reduction in the function and/or numbers of NK cells in most cancers [75, 76]. Investigators found that NK cells were diminished in the EC tumor infiltrate. Tumor resident CD56+CD103+ NK cells expressed higher levels of immune checkpoint inhibitor molecule T cell immunoglobulin and mucin domain-containing protein-3 (TIM-3) (a parameter of exhausted T cells) in comparison with recruited CD56+CD103- NK cells [77]. Further study showed that TIM-3 expression on tumor NK cells from EC patients with lymph node (LN) invasion was higher in these patients with worse disease, than in those

without LN invasion [77]. Differential levels of critical soluble molecules capable of regulating NK cell function were found in healthy and diseased tissue. For example, CCL27, an NK cell chemoattractant molecule, and CCL21 which recruits immune cells to tumors to establish cellular immune responses, were significantly reduced in tumor tissue compared to adjacent healthy tissue. In contrast, the cytokine IL-6, a negative regulator of NK cell function and promoter of tumorigenesis was increased in the tumor. In this same study, cytolytic function was assessed by studying mediators granzyme B, CD107 (degranulation marker) and IFN- γ . Tumor NK cells produced less IFN- γ , TNF- α , granzyme B and CD107, than those of adjacent healthy tissue [77]. These parameters all support the idea of a less efficient NK cell in EC progression.

In patients with stage I EC, NK cell activity was lower with increased nuclear grade [78]. The human leukocyte antigen (HLA) -class 1 system comprises of the HLA-A, -B and -C, as well as the non-classical HLA-E, -F and -G antigens. The receptor NKG2A is selectively expressed on cytotoxic lymphocytes, such as NK cells and CD8+ T cells. NKG2A binds to its ligand HLA-E which is commonly detected in human cancers [79]. In one study, using tissue microarrays (TMAs) of endometrial tumors, investigators reported that upregulation of HLA-E predicted PFS and OS in EC patients. The number of NK cells were associated with survival when HLA-E expression was upregulated, however these cells were associated with worse outcome when HLA-E expression was normal. Thus, in this study, it appears that the prognostic benefit of NK cells is influenced by HLA-E expression in EC [80]. Other cell types which are not well investigated but may be important in EC include cells such as neutrophils. In a study of 510 EC patients, it was determined that increased neutrophil to lymphocyte ratio was associated with advanced stage ($P=0.039$), higher histological grade ($P=0.005$) and lymph node metastasis ($P=0.041$) [81].

Regulation of Endometrial Cancer by Cell Surface Receptors and Soluble Molecules

Immune checkpoint molecule PD-1 (CD279) is predominantly expressed on T cells, and programmed death-1 ligand (PD-L1; CD274) on antigen presenting cells (APC), immunosuppressive macrophages and tumor cells. These molecules are generally overexpressed in tumors. Linkage of PD-1 to PD-L1 is a potent immune-suppressive mechanism in the TME. In cancers, ligation of PD-1 to PD-L1 is generally consistent with a TME abundant in MDSC with increased activity, CD8+ T cells with low cytotoxic potential, and other immunosuppressive parameters which favour tumor progression [82-84].

POLE-ultramutated and MSI-H EC tumors have overexpression of programmed death-1 ligand (PD-L1). Additionally, POLE-ultramutated and MSI/ MMR deficiency (MMRd) tumors generally have high CD3+ and high cytotoxic CD8+ T cells, correlating with the best outcome to immune checkpoint inhibitor therapy of the four EC groups described [21, 23, 85-88]. With the appropriate selection of EC patients, blocking the PD-1/ PD-L1 interaction axis with monotherapy treatment may lead to an ORR as high as 57 % [41]. This is a very successful outcome for an immunotherapy regimen. There are now several FDA approved antibodies such as Pembrolizumab targeting the PD-1 axis, many of

which are in single and combination therapy clinical trials for endometrial cancer and other malignancies [89-91].

In a cohort of 486 EC patients investigators reported the loss of the classical HLA-class 1 (HLA-A and/or -B/-C) molecules was found in 41.3% patients, and more frequently in high grade EC than in low grade cancers [92]. Inhibition of these HLA-class 1 molecules may be an immune escape mechanism whereby cancer cells evade the immune response.

Several soluble molecules are also associated with EC stage or EC prognosis [93]. Levels of interleukin-6 (IL-6, a pro-inflammatory cytokine) mRNA expression and of IL-6 were significantly elevated in uterine serous papillary carcinoma [94].

Indoleamine 2, 3- dioxygenase (IDO) is an enzyme which catalyzes the metabolism of the amino acid tryptophan in the initial stages of the kynurenine pathway [95]. IDO enhances tumor angiogenesis and spread and downregulates tumor infiltrating lymphocyte (TIL) proliferation and other anti-tumor functions. In EC patients, high levels of IDO were found in patients who had shorter PFS and OS, when compared with EC patients with low IDO expression. This parameter of high IDO expression also correlated with low TIL density and low NK cell frequency [96]. Targeting IDO may be a useful strategy for some EC patients.

Cyclooxygenase-2 (COX-2) regulates the synthesis of prostaglandins. In EC patients COX-2 overexpression was positively associated with EC stage [97]. There was an inverse relationship between COX-2 expression and the frequency of CD8+ T cells in tumors. COX-2 levels were a good predictor of EC recurrence [98]. The relevance of COX-2 in EC is further supported by a study which showed that MSI positive EC and high COX-2 expression patients had worse outcome than MSI positive EC patients with low COX-2 expression [99]. The use of COX-2 inhibitors may hold promise as an EC therapy option.

Conclusion

Even though there are a limited number of studies in the literature characterizing the TME of EC patients, it is evident that immune-suppression in EC is multifaceted, and that clues for improved and novel treatments for this disease will result after a better understanding of the immune cells, receptors, soluble molecules and molecular targets in the TME of EC patients. Furthermore, it needs to be determined in preclinical studies which mechanisms of tumor evasion are the most critical to EC progression and poor prognosis so that we can focus primarily on developing treatments which will be most effective.

The modulation of cells of the TME with immunotherapeutic and targeted agents provides a basis for increased use of these agents in combination in clinical practice to improve the dismal survival rate in advanced/metastatic EC. So far, immune checkpoint inhibitors have resulted in good outcome in some subgroups of EC patients. It is however unlikely that any single agent will be highly effective in any large cohort of EC patients. Thus, moving forward, we must consider a combination therapy approach with immune checkpoint inhibitors, vaccines, targeted therapy and conventional treatment agents, to

simultaneously address the diverse nature of immune-suppression in the EC TME. It is hopeful that the outcome of ongoing clinical trials and the design of novel clinical trials with combination treatment regimens in EC patients will significantly advance the field of EC therapy, and soon provide much needed translational options for patients with advanced/metastatic EC, resulting in improved survival.

Funding

None.

Conflicts of Interest

None.

REFERENCES

- Siegel RL, Miller KD, Fuchs HE, Jemal A (2021) Cancer Statistics, 2021. *CA Cancer J Clin* 71: 7-33. [\[Crossref\]](#)
- Yang X, Wang J (2019) The Role of Metabolic Syndrome in Endometrial Cancer: A Review. *Front Oncol* 9: 744. [\[Crossref\]](#)
- Van Arsdale A, Miller DT, Kuo DY, Isani S, Sanchez L et al. (2019) Association of Obesity with Survival in Patients with Endometrial Cancer. *Gynecol Oncol* 154: 156-162. [\[Crossref\]](#)
- Alard E, Butnariu AB, Grillo M, Kirkham C, Zinovkin DA et al. (2020) Advances in Anti-Cancer Immunotherapy: Car-T Cell, Checkpoint Inhibitors, Dendritic Cell Vaccines, and Oncolytic Viruses, and Emerging Cellular and Molecular Targets. *Cancers (Basel)* 12: 1826. [\[Crossref\]](#)
- Bokhman JV (1983) Two Pathogenetic Types of Endometrial Carcinoma. *Gynecol Oncol* 15: 10-17. [\[Crossref\]](#)
- Llobet D, Pallares J, Yeramian A, Santacana M, Eritja N et al. (2009) Molecular Pathology of Endometrial Carcinoma: Practical Aspects from the Diagnostic and Therapeutic Viewpoints. *J Clin Pathol* 62: 777-785. [\[Crossref\]](#)
- Cancer Genome Atlas Research Network, Kandoth C, Schultz N, Cherniack AD, Akbani R et al. (2013) Integrated genomic characterization of endometrial carcinoma. *Nature* 497: 67-73. [\[Crossref\]](#)
- Mittica G, Ghisoni E, Giannone G, Aglietta M, Genta S et al. (2017) Checkpoint Inhibitors in Endometrial Cancer: Preclinical Rationale and Clinical Activity. *Oncotarget* 8: 90532-90544. [\[Crossref\]](#)
- Lynch HT, Snyder CL, Shaw TG, Heinen CD, Hitchins MP (2015) Milestones of Lynch Syndrome: 1895-2015. *Nat Rev Cancer* 15: 181-194. [\[Crossref\]](#)
- Egoavil C, Alenda C, Castillejo A, Paya A, Peiro G et al. (2013) Prevalence of Lynch Syndrome among Patients with Newly Diagnosed Endometrial Cancers. *PLoS One* 8: e79737. [\[Crossref\]](#)
- Haralalddottir S, Hampel H, Tomsic J, Frankel WL, Pearlman R et al. (2014) Colon and Endometrial Cancers with Mismatch Repair Deficiency can Arise from Somatic, rather than Germline, Mutations. *Gastroenterology* 147: 1308.e1-1316.e1. [\[Crossref\]](#)
- Blank CU, Haanen JB, Ribas A, Schumacher TN (2016) CANCER IMMUNOLOGY. The "cancer immunogram". *Science* 352: 658-660. [\[Crossref\]](#)
- Noël G, Langou Fontsa M, Willard Gallo K (2018) The Impact of Tumor Cell Metabolism on T Cell-Mediated Immune Responses and Immuno-Metabolic Biomarkers in Cancer. *Semin Cancer Biol* 52: 66-74. [\[Crossref\]](#)
- Bruno V, Corrado G, Baci D, Chiofalo B, Carosi MA et al. (2020) Endometrial Cancer Immune Escape Mechanisms: Let Us Learn from the Fetal-Maternal Interface. *Front Oncol* 10: 156. [\[Crossref\]](#)
- Hwang W, Adams SF, Tahirovic E, Hagemann IS, Coukos G (2012) Prognostic Significance of Tumor-Infiltrating T Cells in Ovarian Cancer: A Meta-Analysis. *Gynecol Oncol* 124: 192-198. [\[Crossref\]](#)
- Fridman WH, Pagès F, Saut's Fridman C, Galon J (2012) The Immune Contexture in Human Tumours: Impact on Clinical Outcome. *Nat Rev Cancer* 12: 298-306. [\[Crossref\]](#)
- Pascual García M, Bértolo C, Nieto JC, Serrat N, Espinosa Í et al. (2016) CD8 Down-Regulation on Cytotoxic T Lymphocytes of Patients with Endometrioid Endometrial Carcinomas. *Hum Pathol* 56: 180-188. [\[Crossref\]](#)
- Kondratiev S, Sabo E, Yakirevich E, Lavie O, Resnick MB (2004) Intratumoral CD8+ T Lymphocytes as a Prognostic Factor of Survival in Endometrial Carcinoma. *Clin Cancer Res* 10: 4450-4456. [\[Crossref\]](#)
- de Jong RA, Leffers N, Boezen HM, ten Hoor KA, van der Zee AG et al. (2009) Presence of Tumor-Infiltrating Lymphocytes is an Independent Prognostic Factor in Type I and II Endometrial Cancer. *Gynecol Oncol* 114: 105-110. [\[Crossref\]](#)
- Chen W, Hu T, He C (2021) The Prognostic Value of Tumor-Infiltrating Immune Cells in Gynecologic Cancers. *bioRxiv*.
- McConechy MK, Talhouk A, Leung S, Chiu D, Yang W et al. (2016) Endometrial Carcinomas with POLE Exonuclease Domain Mutations have a Favorable Prognosis. *Clin Cancer Res* 22: 2865-2873. [\[Crossref\]](#)
- Billingsley CC, Cohn DE, Mutch DG, Hade EM, Goodfellow PJ (2016) Prognostic Significance of POLE Exonuclease Domain Mutations in High-Grade Endometrioid Endometrial Cancer on Survival and Recurrence: A Subanalysis. *Int J Gynecol Cancer* 26: 933-938. [\[Crossref\]](#)
- Meng B, Hoang LN, McIntyre JB, Duggan MA, Nelson GS et al. (2014) POLE Exonuclease Domain Mutation Predicts Long Progression-Free Survival in Grade 3 Endometrioid Carcinoma of the Endometrium. *Gynecol Oncol* 134: 15-19. [\[Crossref\]](#)
- Guo F, Dong Y, Tan Q, Kong J, Yu B (2020) Tissue Infiltrating Immune Cells as Prognostic Biomarkers in Endometrial Cancer: A Meta-Analysis. *Dis Markers* 2020: 1805764. [\[Crossref\]](#)
- Wakatsuki K, Sho M, Yamato I, Takayama T, Matsumoto S et al. (2013) Clinical Impact of Tumor-Infiltrating CD45RO+ Memory T Cells on Human Gastric Cancer. *Oncol Rep* 29: 1756-1762. [\[Crossref\]](#)
- Pagès F, Berger A, Camus M, Sanchez Cabo F, Costes A et al. (2005) Effector Memory T Cells, Early Metastasis, and Survival in Colorectal Cancer. *N Engl J Med* 353: 2654-2666. [\[Crossref\]](#)
- Wolf D, Wolf AM, Rumpold H, Fiegl H, Zeimet AG et al. (2005) The expression of the regulatory T cell-specific forkhead box transcription factor FoxP3 is associated with poor prognosis in ovarian cancer. *Clin Cancer Res* 11: 8326-8331. [\[Crossref\]](#)
- Zhan L, Liu X, Zhang J, Cao Y, Wei B (2020) Immune Disorder in Endometrial Cancer: Immunosuppressive Microenvironment, Mechanisms of Immune Evasion and Immunotherapy. *Oncol Lett* 20: 2075-2090. [\[Crossref\]](#)
- Chang WC, Li CH, Huang SC, Chang DY, Chou LY et al. (2010) Clinical Significance of Regulatory T Cells and CD8+ Effector

- Populations in Patients with Human Endometrial Carcinoma. *Cancer* 116: 5777-5788. [\[Crossref\]](#)
30. Cao X, Cai SF, Fehniger TA, Song J, Collins LI et al. (2007) Granzyme B and Perforin are Important for Regulatory T Cell-Mediated Suppression of Tumor Clearance. *Immunity* 27: 635-646. [\[Crossref\]](#)
 31. Gondek DC, Lu LF, Quezada SA, Sakaguchi S, Noelle RJ (2005) Cutting Edge: Contact-Mediated Suppression by CD4+CD25+ Regulatory Cells Involves a Granzyme B-Dependent, Perforin-Independent Mechanism. *J Immunol* 174: 1783-1786. [\[Crossref\]](#)
 32. Shang B, Liu Y, Jiang SJ, Liu Y (2015) Prognostic Value of Tumor-Infiltrating FoxP3+ Regulatory T Cells in Cancers: A Systematic Review and Meta-Analysis. *Sci Rep* 5: 15179. [\[Crossref\]](#)
 33. Ma GF, Miao Q, Liu YM, Gao H, Lian JJ et al. (2014) High FoxP3 Expression in Tumour Cells Predicts Better Survival in Gastric Cancer and its Role in Tumour Microenvironment. *Br J Cancer* 110: 1552-1560. [\[Crossref\]](#)
 34. Zhang Y, Lazarus J, Steele NG, Yan W, Lee HJ et al. (2020) Regulatory T-Cell Depletion Alters the Tumor Microenvironment and Accelerates Pancreatic Carcinogenesis. *Cancer Discov* 10: 422-439. [\[Crossref\]](#)
 35. Rowshanravan B, Halliday N, Sansom DM (2018) CTLA-4: a moving target in immunotherapy. *Blood* 131: 58-67. [\[Crossref\]](#)
 36. Miller J, Baker C, Cook K, Graf B, Sanchez Lockhart M et al. (2009) Two Pathways of Costimulation through CD28. *Immunol Res* 45: 159-172. [\[Crossref\]](#)
 37. Friese C, Harbst K, Borch TH, Westergaard MCW, Pedersen M et al. (2020) CTLA-4 Blockade Boosts the Expansion of Tumor-Reactive CD8(+) Tumor-Infiltrating Lymphocytes in Ovarian Cancer. *Sci Rep* 10: 3914. [\[Crossref\]](#)
 38. Sun C, Mezzadra R, Schumacher TN (2018) Regulation and Function of the PD-L1 Checkpoint. *Immunity* 48: 434-452. [\[Crossref\]](#)
 39. Chamoto K, Al Habsi M, Honjo T (2017) Role of PD-1 in Immunity and Diseases. *Curr Top Microbiol Immunol* 410: 75-97. [\[Crossref\]](#)
 40. Marcus L, Lemery SJ, Keegan P, Pazdur R (2019) FDA Approval Summary: Pembrolizumab for the Treatment of Microsatellite Instability-High Solid Tumors. *Clin Cancer Res* 25: 3753-3758. [\[Crossref\]](#)
 41. Marabelle A, Le DT, Ascierto PA, Di Giacomo AM, De Jesus Acosta A et al. (2020) Efficacy of Pembrolizumab in Patients with Noncolorectal High Microsatellite Instability/Mismatch Repair-Deficient Cancer: Results from the Phase II KEYNOTE-158 Study. *J Clin Oncol* 38: 1-10. [\[Crossref\]](#)
 42. Elliott LA, Doherty GA, Sheahan K, Ryan EJ (2017) Human Tumor-Infiltrating Myeloid Cells: Phenotypic and Functional Diversity. *Front Immunol* 8: 86. [\[Crossref\]](#)
 43. Barros MHM, Hauck F, Dreyer JH, Kempkes B, Niedobitek G (2017) Macrophage polarisation: an immunohistochemical approach for identifying M1 and M2 macrophages. *PLoS One* 8: e80908. [\[Crossref\]](#)
 44. Gabrilovich DI (2017) Myeloid-Derived Suppressor Cells. *Cancer Immunol Res* 5: 3-8. [\[Crossref\]](#)
 45. Liu Y, Cao X (2015) The Origin and Function of Tumor-Associated Macrophages. *Cell Mol Immunol* 12: 1-4. [\[Crossref\]](#)
 46. Chen Y, Song Y, Du W, Gong L, Chang H et al. (2019) Tumor-Associated Macrophages: An Accomplice in Solid Tumor Progression. *J Biomed Sci* 26: 78. [\[Crossref\]](#)
 47. Lan C, Huang X, Lin S, Huang H, Cai Q et al. (2013) Expression of M2-Polarized Macrophages is Associated with Poor Prognosis for Advanced Epithelial Ovarian Cancer. *Technol Cancer Res Treat* 12: 259-267. [\[Crossref\]](#)
 48. Qian BZ, Pollard JW (2010) Macrophage Diversity Enhances Tumor Progression and Metastasis. *Cell* 141: 39-51. [\[Crossref\]](#)
 49. Zhan L, Liu X, Zhang J, Cao Y, Wei B (2020) Immune Disorder in Endometrial Cancer: Immunosuppressive Microenvironment, Mechanisms of Immune Evasion and Immunotherapy. *Oncol Lett* 20: 2075-2090. [\[Crossref\]](#)
 50. Dun EC, Hanley K, Wieser F, Bohman S, Yu J et al. (2013) Infiltration of Tumor-Associated Macrophages is Increased in the Epithelial and Stromal Compartments of Endometrial Carcinomas. *Int J Gynecol Pathol* 32: 576-584. [\[Crossref\]](#)
 51. Soeda S, Nakamura N, Ozeki T, Nishiyama H, Hojo H et al. (2008) Tumor-Associated Macrophages Correlate with Vascular Space Invasion and Myometrial Invasion in Endometrial Carcinoma. *Gynecol Oncol* 109: 122-128. [\[Crossref\]](#)
 52. Kübler K, Ayub TH, Weber SK, Zivanovic O, Abramian A et al. (2014) Prognostic significance of tumor-associated macrophages in endometrial adenocarcinoma. *Gynecol Oncol* 135: 176-183. [\[Crossref\]](#)
 53. Talmadge JE, Gabrilovich DI (2013) History of myeloid-derived suppressor cells. *Nat Rev Cancer* 13: 739-752. [\[Crossref\]](#)
 54. Lappat EJ, Cawein M (1964) A Study of the Leukemoid Response to Transplantable A-280 Tumor in Mice. *Cancer Res* 24: 302-311. [\[Crossref\]](#)
 55. De Cicco P, Ercolano G, Ianaro A (2020) The New Era of Cancer Immunotherapy: Targeting Myeloid-Derived Suppressor Cells to Overcome Immune Evasion. *Front Immunol* 11: 1680. [\[Crossref\]](#)
 56. Kumar V, Patel S, Tcyganov E, Gabrilovich DI (2016) The Nature of Myeloid-Derived Suppressor Cells in the Tumor Microenvironment. *Trends Immunol* 37: 208-220. [\[Crossref\]](#)
 57. Dysthe M, Parihar R (2020) Myeloid-Derived Suppressor Cells in the Tumor Microenvironment. *Adv Exp Med Biol* 1224: 117-140. [\[Crossref\]](#)
 58. Safarzadeh E, Orangi M, Mohammadi H, Babaie F, Baradaran B (2018) Myeloid-Derived Suppressor Cells: Important Contributors to Tumor Progression and Metastasis. *J Cell Physiol* 233: 3024-3036. [\[Crossref\]](#)
 59. Meyer C, Cagnon L, Costa Nunes CM, Baumgaertner P, Montandon N et al. (2014) Frequencies of Circulating MDSC Correlate with Clinical Outcome of Melanoma Patients Treated with Ipilimumab. *Cancer Immunol Immunother* 63: 247-257. [\[Crossref\]](#)
 60. Wang Y, Jia A, Bi Y, Wang Y, Yang Q et al. (2020) Targeting Myeloid-Derived Suppressor Cells in Cancer Immunotherapy. *Cancers (Basel)* 12: 2626. [\[Crossref\]](#)
 61. Mandruzzato S, Brandau S, Britten CM, Bronte V, Damuzzo V et al. (2016) Toward Harmonized Phenotyping of Human Myeloid-Derived Suppressor Cells by Flow Cytometry: Results from an Interim Study. *Cancer Immunol Immunother* 65: 161-169. [\[Crossref\]](#)
 62. Rodriguez PC, Quiceno DG, Zabaleta J, Ortiz B, Zea AH et al. (2004) Arginase I Production in the Tumor Microenvironment by Mature Myeloid Cells Inhibits T-Cell Receptor Expression and Antigen-Specific T-Cell Responses. *Cancer Res* 64: 5839-5849. [\[Crossref\]](#)
 63. Stiff A, Trikha P, Mundy Bosse B, McMichael E, Mace TA et al. (2018) Nitric Oxide Production by Myeloid-Derived Suppressor Cells Plays a Role in Impairing Fc Receptor-Mediated Natural Killer Cell Function. *Clin Cancer Res* 24: 1891-1904. [\[Crossref\]](#)
 64. Trovato R, Fiore A, Sartori S, Canè S, Giugno R et al. (2019) Immunosuppression by monocytic myeloid-derived suppressor cells in

- patients with pancreatic ductal carcinoma is orchestrated by STAT3. *J Immunother Cancer* 7: 255. [\[Crossref\]](#)
65. de Aguiar RB, de Moraes JZ (2019) Exploring the Immunological Mechanisms Underlying the Anti-Vascular Endothelial Growth Factor Activity in Tumors. *Front Immunol* 10: 1023. [\[Crossref\]](#)
 66. Anfray C, Ummano A, Andón FT, Allavena P (2019) Current Strategies to Target Tumor-Associated-Macrophages to Improve Anti-Tumor Immune Responses. *Cells* 9: 46. [\[Crossref\]](#)
 67. Banchereau J, Briere F, Caux C, Davoust J, Lebecque S et al. (2000) Immunobiology of dendritic cells. *Annu Rev Immunol* 18: 767-811. [\[Crossref\]](#)
 68. Strioga M, Schijns V, Powell DJ, Pasukoniene V, Dobrovolskiene N et al. (2013) Dendritic cells and their role in tumor immunosurveillance. *Innate Immun* 19: 98-111. [\[Crossref\]](#)
 69. Solano Gálvez SG, Tovar Torres SM, Tron Gómez MS, Weiser Smeke AE, Álvarez Hernández DA et al. (2018) Human Dendritic Cells: Ontogeny and their Subsets in Health and Disease. *Med Sci (Basel)* 6: 88. [\[Crossref\]](#)
 70. Veglia F, Gabrilovich DI (2017) Dendritic Cells in Cancer: The Role Revisited. *Curr Opin Immunol* 45: 43-51. [\[Crossref\]](#)
 71. Wylie B, Macri C, Mintern JD, Waithman J (2019) Dendritic Cells and Cancer: From Biology to Therapeutic Intervention. *Cancers (Basel)* 11: 521. [\[Crossref\]](#)
 72. Lijun Z, Xin Z, Danhua S, Xiaoping L, Jianliu W et al. (2012) Tumor-Infiltrating Dendritic Cells may be used as Clinicopathologic Prognostic Factors in Endometrial Carcinoma. *Int J Gynecol Cancer* 22: 836-841. [\[Crossref\]](#)
 73. Jia J, Wang Z, Li X, Wang Z, Wang X (2012) Morphological Characteristics and Co-Stimulatory Molecule (CD80, CD86, CD40) Expression in Tumor Infiltrating Dendritic Cells in Human Endometrioid Adenocarcinoma. *Eur J Obstet Gynecol Reprod Biol* 160: 223-227. [\[Crossref\]](#)
 74. van Gulijk M, Dammeijer F, Aerts JGJV, Vroman H (2018) Combination Strategies to Optimize Efficacy of Dendritic Cell-Based Immunotherapy. *Front Immunol* 9: 2759. [\[Crossref\]](#)
 75. Aktaş ON, Öztürk AB, Erman B, Erus S, Tanju S et al. (2018) Role of Natural Killer Cells in Lung Cancer. *J Cancer Res Clin Oncol* 144: 997-1003. [\[Crossref\]](#)
 76. Rocca YS, Roberti MP, Juliá EP, Pampena MB, Bruno L et al. (2016) Phenotypic and Functional Dysregulated Blood NK Cells in Colorectal Cancer Patients can be Activated by Cetuximab Plus IL-2 or IL-15. *Front Immunol* 7: 413. [\[Crossref\]](#)
 77. Degos C, Heinemann M, Barrou J, Boucherit N, Lambaudie E et al. (2019) Endometrial Tumor Microenvironment Alters Human NK Cell Recruitment, and Resident NK Cell Phenotype and Function. *Front Immunol* 10: 877. [\[Crossref\]](#)
 78. Garzetti GG, Ciavattini A, Goteri G, Tranquilli AL, Muzzioli M et al. (1994) Natural Killer Cell Activity in Stage I Endometrial Carcinoma: Correlation with Nuclear Grading, Myometrial Invasion, and Immunoreactivity of Proliferating Cell Nuclear Antigen. *Gynecol Oncol* 55: 111-114. [\[Crossref\]](#)
 79. Borst L, van der Burg SH, van Hall T (2020) The NKG2A-HLA-E Axis as a Novel Checkpoint in the Tumor Microenvironment. *Clin Cancer Res* 26: 5549-5556. [\[Crossref\]](#)
 80. Versluis MAC, Marchal S, Plat A, de Bock GH, van Hall T et al. (2017) The prognostic benefit of tumour-infiltrating Natural Killer cells in endometrial cancer is dependent on concurrent overexpression of Human Leucocyte Antigen-E in the tumour microenvironment. *Eur J Cancer* 86: 285-295. [\[Crossref\]](#)
 81. Dong Y, Cheng Y, Wang J (2019) The Ratio of Neutrophil to Lymphocyte is a Predictor in Endometrial Cancer. *Open Life Sci* 14: 110-118.
 82. Alsaab HO, Sau S, Alzhrani R, Tatiparti K, Bhise K et al. (2017) PD-1 and PD-L1 Checkpoint Signaling Inhibition for Cancer Immunotherapy: Mechanism, Combinations, and Clinical Outcome. *Front Pharmacol* 8: 561. [\[Crossref\]](#)
 83. Ahmadzadeh M, Johnson LA, Heemsker B, Wunderlich JR, Dudley ME et al. (2009) Tumor Antigen-Specific CD8 T Cells Infiltrating the Tumor Express High Levels of PD-1 and are Functionally Impaired. *Blood* 114: 1537-1544. [\[Crossref\]](#)
 84. Iwai Y, Hamanishi J, Chamoto K, Honjo T (2017) Cancer Immunotherapies Targeting the PD-1 Signaling Pathway. *J Biomed Sci* 24: 26. [\[Crossref\]](#)
 85. Dudley JC, Lin MT, Le DT, Eshleman JR (2016) Microsatellite Instability as a Biomarker for PD-1 Blockade. *Clin Cancer Res* 22: 813-820. [\[Crossref\]](#)
 86. Zhang S, Minaguchi T, Xu C, Qi N, Itagaki H et al. (2020) PD-L1 and CD4 are Independent Prognostic Factors for overall Survival in Endometrial Carcinomas. *BMC Cancer* 20: 127. [\[Crossref\]](#)
 87. Yamashita H, Nakayama K, Ishikawa M, Nakamura K, Ishibashi T et al. (2017) Microsatellite Instability is a Biomarker for Immune Checkpoint Inhibitors in Endometrial Cancer. *Oncotarget* 9: 5652-5664. [\[Crossref\]](#)
 88. Howitt BE, Shukla SA, Sholl LM, Ritterhouse LL, Watkins JC et al. (2015) Association of Polymerase E-Mutated and Microsatellite-Instable Endometrial Cancers with Neoantigen Load, Number of Tumor-Infiltrating Lymphocytes, and Expression of PD-1 and PD-L1. *JAMA Oncol* 1: 1319-1323. [\[Crossref\]](#)
 89. Marinelli O, Annibali D, Aguzzi C, Tuyaeys S, Amant F et al. (2019) The Controversial Role of PD-1 and its Ligands in Gynecological Malignancies. *Front Oncol* 9: 1073. [\[Crossref\]](#)
 90. Castellano T, Moore KN, Holman LL (2018) An Overview of Immune Checkpoint Inhibitors in Gynecologic Cancers. *Clin Ther* 40: 372-388. [\[Crossref\]](#)
 91. Vaddepally RK, Kharel P, Pandey R, Garje R, Chandra AB (2020) Review of Indications of FDA-Approved Immune Checkpoint Inhibitors Per NCCN Guidelines with the Level of Evidence. *Cancers (Basel)* 12: 738. [\[Crossref\]](#)
 92. de Jong RA, Boerma A, Boezen HM, Mourits MJ, Hollema H et al. (2012) Loss of HLA Class I and Mismatch Repair Protein Expression in Sporadic Endometrioid Endometrial Carcinomas. *Int J Cancer* 131: 1828-1836. [\[Crossref\]](#)
 93. Subramaniam KS, Omar IS, Kwong SC, Mohamed Z, Woo YL et al. (2016) Cancer-Associated Fibroblasts Promote Endometrial Cancer Growth Via Activation of Interleukin-6/STAT-3/C-Myc Pathway. *Am J Cancer Res* 6: 200-213. [\[Crossref\]](#)
 94. Bellone S, Watts K, Cane S, Palmieri M, Cannon MJ et al. (2005) High serum levels of interleukin-6 in endometrial carcinoma are associated with uterine serous papillary histology, a highly aggressive and chemotherapy-resistant variant of endometrial cancer. *Gynecol Oncol* 98: 92-98. [\[Crossref\]](#)
 95. Ricciuti B, Leonardi GC, Puccetti P, Fallarino F, Bianconi V et al. (2019) Targeting Indoleamine-2,3-Dioxygenase in Cancer: Scientific

- Rationale and Clinical Evidence. *Pharmacol Ther* 196: 105-116. [[Crossref](#)]
96. Yu CP, Fu SF, Chen X, Ye J, Ye Y et al. (2018) The Clinicopathological and Prognostic Significance of IDO1 Expression in Human Solid Tumors: Evidence from a Systematic Review and Meta-Analysis. *Cell Physiol Biochem* 49: 134-143. [[Crossref](#)]
97. Li M, Li M, Wei Y, Xu H (2020) Prognostic and Clinical Significance of Cyclooxygenase-2 Overexpression in Endometrial Cancer: A Meta-Analysis. *Front Oncol* 10: 1202. [[Crossref](#)]
98. Ohno Y, Ohno S, Suzuki N, Kamei T, Inagawa H et al. (2005) Role of Cyclooxygenase-2 in Immunomodulation and Prognosis of Endometrial Carcinoma. *Int J Cancer* 114: 696-701. [[Crossref](#)]
99. Suemori T, Susumu N, Iwata T, Banno K, Yamagami W et al. (2015) Intratumoral CD8+ Lymphocyte Infiltration as a Prognostic Factor and its Relationship with Cyclooxygenase 2 Expression and Microsatellite Instability in Endometrial Cancer. *Int J Gynecol Cancer* 25: 1165-1172. [[Crossref](#)]